

Olfactomedin 4 Inhibits Cathepsin C-Mediated Protease Activities, Thereby Modulating Neutrophil Killing of *Staphylococcus aureus* and *Escherichia coli* in Mice

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Neutrophils kill bacteria generally through oxidative and nonoxidative mechanisms. Whereas much research has focused on the enzymes essential for neutrophil killing, little is known about the regulatory molecules responsible for such killing. In this study, we investigated the role of olfactomedin 4 (OLFM4), an olfactomedin-related glycoprotein, in neutrophil bactericidal capability and host innate immunity. Neutrophils from *OLFM4*^{-/-} mice have increased intracellular killing of *Staphylococcus aureus* and *Escherichia coli* in vitro. The *OLFM4*^{-/-} mice have enhanced in vivo bacterial clearance and are more resistant to sepsis when challenged with *S. aureus* or *E. coli* by i.p. injection. OLFM4 was found to interact with cathepsin C, a cysteine protease that plays an important role in bacterial killing and immune regulation. We demonstrated that OLFM4 inhibited cathepsin C activity in vitro and in vivo. The cathepsin C activity in neutrophils from *OLFM4*^{-/-} mice was significantly higher than that in neutrophils from wild-type littermate mice. The activities of three serine proteases (neutrophil elastase, cathepsin G, and proteinase 3), which require cathepsin C activity for processing and maturity, were also significantly higher in *OLFM4*^{-/-} neutrophils. The bacterial killing and clearance capabilities observed in *OLFM4*^{-/-} mice that were enhanced relative to wild-type mice were significantly compromised by the additional loss of cathepsin C in mice with OLFM4 and cathepsin C double deficiency. These results indicate that OLFM4 is an important negative regulator of neutrophil bactericidal activity by restricting cathepsin C activity and its downstream granule-associated serine proteases. *The Journal of Immunology*, 2012, 189: 2460–2467.

Neutrophils are a major component of innate immunity, providing a crucial first barrier against bacterial infection. It has been well documented that NADPH-mediated oxidative burst acts to kill certain pathogens (1). Other than reactive oxygen species, neutrophils also produce a multitude of antimicrobial molecules in their granules. The granules fuse with the phagolysosomes and contain defensins, bactericidal permeability-increasing protein, as well as serine proteases such as elastase, cathepsin G, and proteinase 3. The serine proteases generally degrade bacterial proteins and are major antimicrobial components. It has been shown that serine proteases, when activated, are primarily responsible for the destruction of bacteria (1). Recent studies suggest that bacteria are not killed by a common mechanism (2).

Cathepsin C (also called dipeptidyl peptidase I) is a lysosomal cysteine peptidase belonging to the papain family (3) and has numerous biological functions. In addition to a proposed degradative role as a lysosomal exopeptidase, cathepsin C is essential for activating granule-associated serine protease in neutrophils (elastase, cathepsin G, and proteinase 3) (4). These serine proteases require the activity cathepsin C to remove prodipeptide to become active. Serine proteases are mainly stored in azurophilic granules in their active forms until they are released after neutrophil exposure to active stimuli and have broad biological effects including intracellular microbial killing and modulation of inflammatory-cell recruitment (4).

OLFM4 (also called hGC-1 and GW112) is a member of the olfactomedin family, which is characterized by the presence of an olfactomedin domain in the C terminus (5). Olfactomedin-related proteins have distinct tissue expression patterns and diverse cellular functions (5). OLFM4 was first cloned from G-CSF-stimulated human myeloid precursor cells (6). Although most of the other members are preferentially expressed in nervous tissues, OLFM4 is mainly expressed in hematopoietic myeloid cells and in the gastrointestinal tract (6). OLFM4 is involved in regulation of multiple cellular functions including apoptosis (7, 8), differentiation (8), and growth (9). OLFM4 expression is upregulated in some inflammatory diseases, such as chronic inflammatory bowel diseases (10), and in *Helicobacter pylori*-infected patients (11). It was recently reported that OLFM4 downregulates mouse host innate immunity to *H. pylori* infection (12). However, little is known about the physiological functions of phylogenetically conserved OLFM4 in host defense against a broader range of bacteria. Moreover, its expression, localization, and potential biological functions in mature myeloid cells (neutrophils) remain elusive. In this study, we investigated whether OLFM4 regulates host defense against a broader range of bacteria including Gram-positive *Staphylococ-*

cus aureus and Gram-negative *Escherichia coli* using an *OLFM4*^{-/-} mouse model.

Materials and Methods

Mice, cells, and bacteria

OLFM4^{-/-} mice have been described previously (12) and backcrossed six generations into a C57BL/6 background. *Cathepsin C* target mutant mice (*CatC*^{-/-}) in the C57BL/6 background were provided from Washington University School of Medicine (St. Louis, MO). Animals were kept in a specific pathogen-free facility at the National Institutes of Health (Bethesda, MD) and used according to protocols approved by the National Heart, Lung, and Blood Institute Animal Care and Use Committee. All experiments were conducted with animals 10–12 wk of age. HEK 293T cells were purchased from American Type Culture Collection (ATCC; Manassas, VA) and maintained as previously described (13). *S. aureus* ATCC 10390 and *E. coli* ATCC 10536 were purchased from ATCC. *Salmonella enterica* was obtained from Dr. William Coleman's laboratory (National Institute of Diabetes and Digestive and Kidney Disease/National Institutes of Health, Bethesda, MD). All bacteria were grown in tryptic soy broth or on tryptic soy agar plates (Teknova, Hollister, CA).

Yeast two-hybrid library screening

Full-length human *OLFM4* cDNA (bait) was cloned into pCWX200 and pLexA expression plasmid. This bait plasmid was transformed into yeast strain Y304 and screened using a TetR system against a HeLa cDNA library in pYESTrp2 containing 1×10^7 independent clones in strain EGY42. The positive colonies (those appearing blue) were retested for phenotypes. The plasmids from positive colonies were then isolated after transformation into *E. coli*, and the interactions were confirmed in yeast by mating. Plasmids from verified colonies were sequenced.

Isolation of neutrophils

Human peripheral blood neutrophils were isolated from healthy donors using HetaSep (Stem Cell Technologies, Vancouver, BC, Canada) sedimentation and Ficoll-Paque PLUS density gradient separation followed by hypotonic lysis of erythrocytes. Human fresh bone-marrow neutrophils were obtained from AllCells (Emeryville, CA). Mouse neutrophils were isolated from bone marrow. The femur was removed from mice, then flushed with PBS containing 0.5% BSA. The mouse bone marrow neutrophils were purified through a three-layer gradient of 78, 69, and 52% Percoll (Amersham Pharmacia Biotech, Uppsala, Sweden) as previously described (14). Mouse bone marrow-derived neutrophils were used in our study based on the finding that these neutrophils are in a resting state and are functionally competent (14).

Subcellular fractionation of neutrophil granules

Neutrophil granules were enriched by centrifugation on a three-layer Percoll density gradient as previously described (15).

Immunoprecipitation and Western blotting

Immunoprecipitation and Western blot analysis were performed as previously described (13). *OLFM4* polyclonal Ab (16) and cathepsin C Ab (Santa Cruz Biotechnology, Santa Cruz, CA) were used as primary Abs (1:1000). HRP-conjugated donkey anti-rabbit or sheep anti-mouse Abs (1:4000; Amersham, Piscataway, NJ) were used as secondary Abs.

Quantitative reverse transcriptase PCR

Total RNA preparation and quantitative reverse transcriptase PCR was performed as previously described (8).

Bactericidal activity

Mouse neutrophils were isolated from the bone marrow. In a 24-well plate, neutrophils were resuspended in DMEM (10^6 cell/ml) and seeded in wells for 30 min. Nonadherent cells were removed by gentle rinsing. Bacteria were preopsonized with mouse serum for 30 min at 37°C. Bacteria (10^7) were added onto the monolayer of cells. After incubation at 37°C for 15 min, the media were gently removed, and neutrophils were lysed in PBS containing 0.1% Triton X-100 after incubation for 30 min in the presence or absence of gentamicin (100 µg/ml) or lysostaphin (50 µg/ml). The number of viable bacteria was determined using the standard plate method (17).

Peritoneum bacterial clearance and neutrophil influx

Mice were injected i.p. with a sublethal dose of *E. coli* (5×10^3) or *S. aureus* (5×10^4). Peritoneal cavities were lavaged with 5 ml PBS 2 h

after inoculation. The number of viable bacteria was determined using the standard plate method. The percentages of blood granulocytes and mononuclear cells were determined by differential counting on cytospin preparations. The number of neutrophils was determined.

Survival and bacterial dissemination post-i.p. infection

E. coli and *S. aureus* were grown in tryptic soy broth to the exponential phase. Mice of each genotype between 10 and 12 wk of age were injected i.p. with varying amounts of bacteria (CFU). An optimal concentration to induce sepsis and mortality within 72 h was determined for *E. coli* (1.5×10^8 CFU) and *S. aureus* (3.5×10^8 CFU). In survival experiments using this concentration, survival was monitored every 6 h. To determine the bacterial dissemination to large organs during sepsis, liver and lung tissues were harvested 24 h after *S. aureus* or *E. coli* infection. The tissues were homogenized with PBS, and aliquots of serially diluted homogenate were plated on tryptic soy agar. The colonies were enumerated the following day.

OLFM4 protein purification

Human *OLFM4* proteins were purified using the LEXSY system (Jena Bioscience, Jena, Germany) from the protozoan host *Leishmania tarentolae*. The *OLFM4* ORF was PCR-amplified from pcDNA3.1/*OLFM4*-V5 vector (13) and inserted into the inducible LEXSY expression vector pLEXSY_I-neo2 in-frame to the LEXSY signal peptide and C-terminal HexaHis. The LEXSY host T7-TR was transfected with an expression plasmid for inducible secretory expression. Human *OLFM4* proteins were purified by NiNTA affinity chromatography from LEXSY culture supernatants. Elutes were dialyzed into TBS (50 mM Tris pH 7.5, 150 mM NaCl).

Protease activity assays

Bone marrow-derived neutrophils were washed twice with PBS and lysed with cathepsin C lysis buffer (25 mM MES, pH 6.0, 50 mM NaCl, 5 mM DTT, 0.2% Nonidet P-40, for cathepsin C assays) or with serine protease lysis buffer (20 mM Tris-HCl, pH 7.5, 100 mM NaCl, 0.2% Nonidet P-40, for serine protease assays). Debris was removed by centrifugation at $15,000 \times g$ for 10 min, and the supernatants were retained. Cathepsin C activities were assayed in 25 mM MES, pH 6.0, 50 mM NaCl, 5 mM DTT, 0.1% PEG 3350 using Gly-Arg-AMC (Bachem, Torrance, CA) at 10 µM. Reaction progress was monitored continuously with product (AMC) on a FLUOstar Optima Fluorimeter (BMG, Gary, NC) with 380-nm excitation and 460-nm emission wavelength filters. Serine protease activities were assayed in 100 mM Tris-HCl, pH 7.5, and 50 mM NaCl supplemented with a peptide substrate specific for the serine protease assessed: for neutrophil elastase, 600 µM methoxysuccinyl-Ala-Ala-Pro-Val-*p*-nitroanilide (Elastin Products Company, Owensville, MO); for cathepsin G, 600 µM N-succinyl-Ala-Ala-Pro-Phe-*p*-nitroanilide (Sigma, St. Louis, MO); and for proteinase 3, 100 µM Boc-Ala-Ala-Nva-SBzL (Elastin Products Company). Release of *p*-nitroanilide was measured by absorbance at 410 nm. Release of SBzL was measured by absorbance at 320 nm. Background was subtracted. For in vitro cathepsin C activity assays, recombinant human cathepsin C (R&D Systems, Minneapolis, MN) was activated by cathepsin L (R&D Systems) in the activation buffer (25 mM MES, 5 mM DTT, pH 6.0) at room temperature for 1 h. Various amounts of purified *OLFM4* proteins were incubated with activated cathepsin C in the activation buffer (25 mM MES, 50 mM NaCl, 5 mM DTT, pH 6.0) for 30 min at room temperature. Cathepsin C activity was measured as described earlier.

Oxidative burst assay

Superoxide production was determined using the Diogenes Cellular Luminescence Enhancement System for Superoxide Detection according to the manufacturer's instructions (National Diagnostics, Atlanta, GA).

Myeloperoxidase activity assay

Myeloperoxidase (MPO) activity was quantitatively measured by the method of Suzuki et al. (18) with some modifications. Neutrophils were adjusted to 5×10^6 cells/ml and incubated with N-formyl-Met-Leu-Phe (1 µM) and cytochalasin B (5 µg/ml) for 10 min at 37°C. Triton X-100 (final concentration, 0.1%) was added to the cell suspensions for total MPO release. Aliquots of the cell extracts were incubated with the TMB liquid substrate system, and the activity was detected spectrophotometrically at 370 nm.

Phagocytosis

Phagocytosis was assessed by flow cytometry as previously described (19). FITC-labeled *S. aureus* or *E. coli* particles (Molecular Probes, Eugene, OR) were reconstituted and opsonized with opsonizing reagent (Molecular Probes) according to the manufacturer's instructions. Opsonized particles

(10^7) were washed and incubated with neutrophils (10^6) (multiplicity of infection = 10). The threshold (forward-scattered light) was set to exclude particles not associated with neutrophils. After measuring the total number of neutrophils with bound/ingested particles, samples were quenched with 0.4% trypan blue to determine the percentage of neutrophils with ingested particles. The percentage phagocytosis was determined by the percentage of FITC⁺ neutrophils observed after quenching.

Statistical analysis

Significance of differences between experimental groups was determined by a two-way Student *t* test. Differences were considered significant when $p < 0.05$. Survival statistics were performed with the Kaplan–Meier log-rank test (GraphPad Prism version 4.0).

Results

OLFM4 is a neutrophil granule protein that responds to bacteria infection

OLFM4 is a secreted glycoprotein (7) and is also retained inside neutrophils (8, 13). We first determined whether OLFM4 is a neutrophil granule protein. Neutrophil granule fractions were separated from human peripheral blood neutrophils using Percoll density gradient (15). OLFM4 protein expression was detected in all three neutrophil granule subsets, with high abundance in specific granules, modest levels in gelatinase and azurophil granules, and absence in secretory vesicles (Fig. 1A). Previously, it has been shown that OLFM4 is upregulated under inflammatory (10) and *H. pylori* infection (11) conditions. In this article, we demonstrated that OLFM4 mRNA expression in human neutrophils was upregulated in response to a broad range of bacterial infections, including Gram-positive *S. aureus* and Gram-negative *E. coli* and *S. enterica* infections (Fig. 1B). These results indicate that OLFM4 is a novel neutrophil granule protein that exhibits enhanced expression to a broad range of bacterial infections.

OLFM4 deficiency in neutrophils leads to an increased bactericidal capacity, enhanced in vivo bacterial clearance, and resistance to sepsis

To determine the mode of action of OLFM4 in combating bacterial infections, we compared the ability of neutrophils from *OLFM4*^{-/-} mice and their wild-type (WT) littermates to recognize and kill bacteria in vitro. We exposed monolayers of neutrophils to *S. aureus* or *E. coli*. After 15 min, unbound bacteria were gently removed, and the number of viable bacteria associated with the neutrophil monolayer was assessed. The number of viable bacteria for *OLFM4*^{-/-} and WT neutrophil groups at 15 min was comparable (Supplemental Fig. 1A). This observation suggests that *OLFM4*^{-/-} and WT neutrophils were able to recognize and bind bacteria equally well. After incubation for an additional 30 min in the presence of gentamicin or lysostaphin that eliminates extracellular *E. coli* or *S. aureus*, respectively, the intracellular bactericidal activity was assessed. Under these conditions, *OLFM4*^{-/-} neu-

trophils exhibited more bactericidal activity than WT neutrophils (Fig. 2A).

We next determined the capability of *OLFM4*^{-/-} and WT mice to clear bacteria from the site of infection in vivo. Mice were challenged i.p. with *S. aureus* or *E. coli*. After 2 h, the number of viable bacteria in the peritoneal cavities of *OLFM4*^{-/-} mice was significantly less than that in WT mice (Fig. 2B). Neutrophil recruitment into the peritoneal cavity was comparable for *OLFM4*^{-/-} and WT mice as determined by neutrophil counts and MPO activity (Supplemental Fig. 1B, 1C).

Furthermore, we compared the susceptibility of *OLFM4*^{-/-} and WT mice to sepsis induced by i.p. challenge with *S. aureus* or *E. coli*. We first determined an optimal bacterial concentration to induce lethal sepsis over 72 h in WT mice, then used this concentration to examine survival. *OLFM4*^{-/-} mice were found to be much more resistant to sepsis than WT mice (Fig. 2C). Consistent with the survival assay, the number of bacteria disseminated to the liver and lung in *OLFM4*^{-/-} mice was significantly lower than that in WT mice postinfection for 24 h (Fig. 2D). These data suggested that endogenous OLFM4 is a critical negative regulator of intracellular bactericidal activity in neutrophils and thus dampens mice host immune response to bacterial infection.

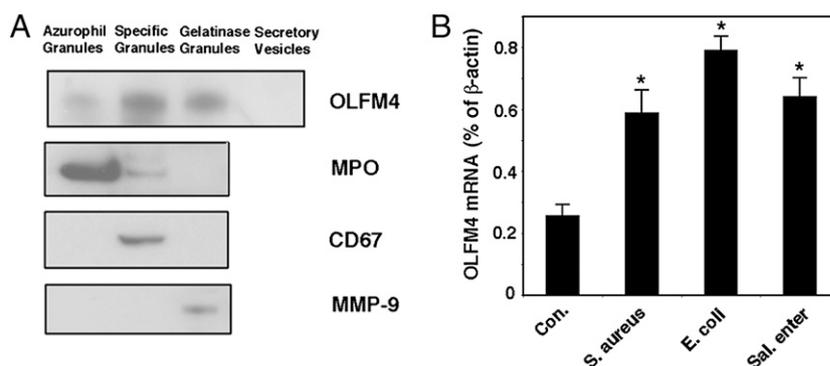
The NADPH oxidative burst, MPO activity, and phagocytosis are not affected in OLFM4^{-/-} neutrophils

The NADPH oxidase-mediated superoxide (O₂⁻) burst (Supplemental Fig. 2), MPO activity (Supplemental Fig. 3), and phagocytosis of *S. aureus* and *E. coli* (Supplemental Fig. 4) were found to be comparable in *OLFM4*^{-/-} and WT neutrophils. We also found that purified recombinant human OLFM4 protein had no direct effect on the growth of *E. coli* and *S. aureus* in vitro (data not shown). These data suggest that OLFM4 regulation of bactericidal activity is not mediated directly through the oxidative pathway, and OLFM4 protein does not have a direct effect on bacteria growth.

OLFM4 binds cathepsin C and inhibits cathepsin C activity in vitro and in vivo

To further investigate the molecular mechanism of OLFM4 in regulating neutrophil killing of bacteria, we performed a yeast two-hybrid assay to identify the potential binding partner of human OLFM4. Cathepsin C, a cysteine protease, was shown to interact with OLFM4 (Fig. 3A). A coimmunoprecipitation assay confirmed that OLFM4 binds to cathepsin C in HEK 293T cells that overexpress full-length OLFM4 and cathepsin C (Fig. 3B). A reciprocal coimmunoprecipitation assay demonstrated endogenous association of OLFM4 with cathepsin C in human bone marrow neutrophils (Fig. 3C). The physical association between OLFM4 and cathepsin C raised the possibility that OLFM4 plays a role in modulating cathepsin C activity. Expression of OLFM4 in HEK 293T cells

FIGURE 1. OLFM4 is a neutrophil granule protein that responds to bacterial infection. (A) Western blotting was performed to detect OLFM4, MPO, CD67, and MMP-9 in human peripheral blood neutrophil granule subsets. (B) Quantitative reverse transcriptase PCR was performed to detect OLFM4 mRNA expression in human peripheral blood neutrophils incubated in vitro in the presence or absence of *S. aureus*, *E. coli*, or *S. enterica* (*Sal. enter*) infection for 6 h (neutrophil/bacteria = 1:10). Data represent expression relative to β -actin expression and are expressed as mean \pm SD ($n = 3$). * $p < 0.05$ when compared with control (Con.).



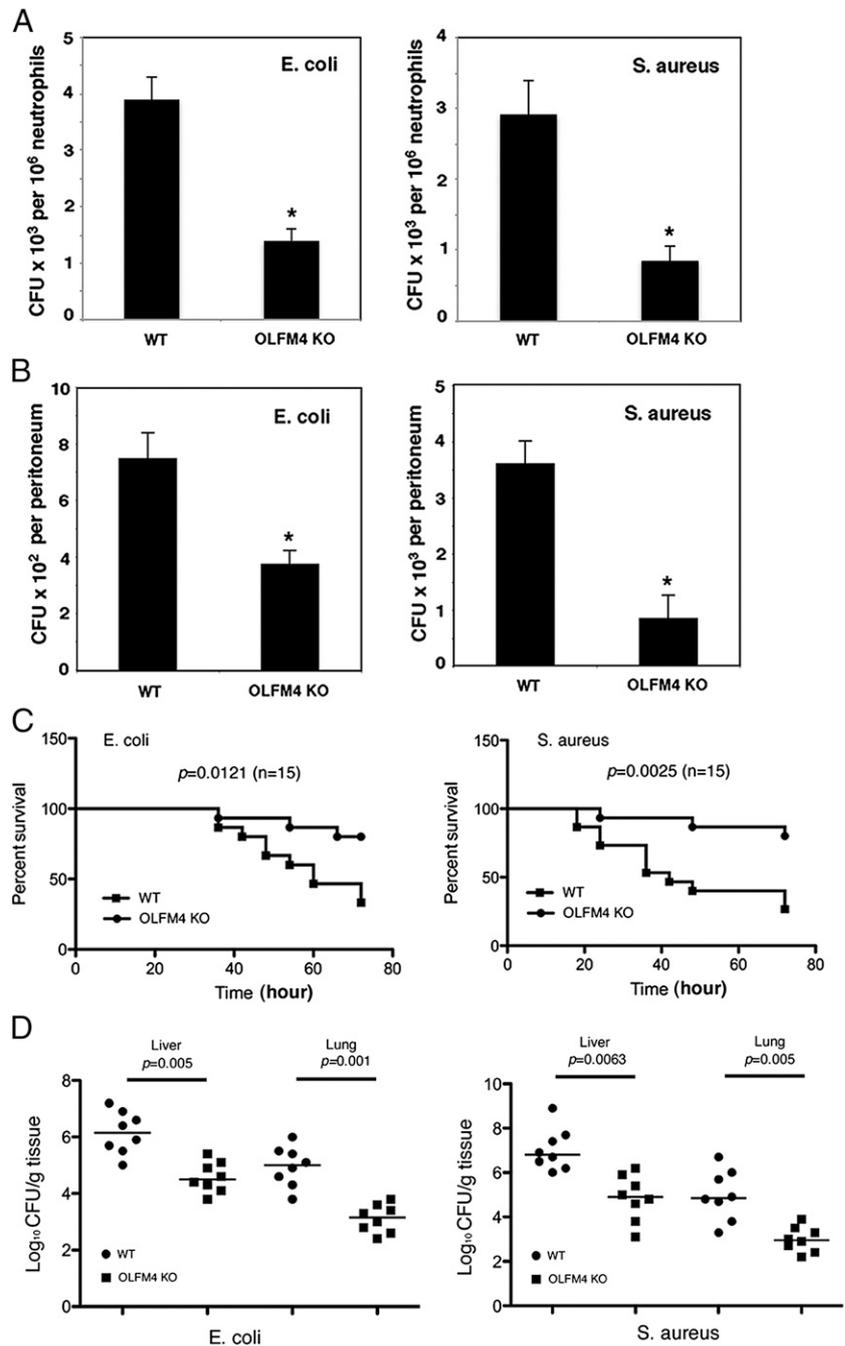


FIGURE 2. *OLFM4*^{-/-} mice display enhanced intracellular bacterial killing, increased peritoneal bacterial clearance, and resistance to sepsis. **(A)** Neutrophils (10⁶) derived from the bone marrow of WT and *OLFM4*^{-/-} (*OLFM4* KO) mice were incubated with preopsonized *E. coli* (10⁷) or *S. aureus* (10⁷) in a 24-well plate. After 15 min, the media were gently removed. The cells were treated with gentamicin (100 μg/ml) for *E. coli* or lysostaphin (50 μg/ml) for *S. aureus* for additional 30 min. The viable bacteria were counted on tryptic soy agar plates. Data are expressed as mean ± SD (*n* = 4). **p* < 0.05. **(B)** Mice were challenged i.p. with *E. coli* (5 × 10³) or *S. aureus* (5 × 10⁴). After 2 h, the peritoneal cavity was lavaged and the number of bacterial colonies was determined on plates. Data are expressed as mean ± SD (*n* = 4). **p* < 0.05. **(C)** Mice (10–12 wk old, *n* = 15) were infected i.p. with *E. coli* (1.5 × 10⁸ CFU) or *S. aureus* (3.5 × 10⁸ CFU), and survival was monitored every 6 h over 72 h. **(D)** Mice (*n* = 8) were treated as in (C). Mice were euthanized 24 h after bacterial infection, and liver and lung tissues were collected. Homogenized tissues were serially diluted and plated, and colonies were enumerated the following day.

suppressed the activity of endogenous cathepsin C and of over-expressed cathepsin C activity (Fig. 3D). Purified OLFM4 protein effectively inhibited recombinant human cathepsin C activity in vitro (Fig. 3E), with an IC₅₀ of 12.5 nM. These results indicate that OLFM4 is an effective inhibitor of cathepsin C in vitro and in vivo.

Cathepsin C and serine protease activities are enhanced in neutrophils of *OLFM4*^{-/-} mice

It has been recently reported that neutrophil serine proteases play much more important roles in killing bacteria than previously thought (1). The processing and activation of serine proteases require cathepsin C activity (4). Having demonstrated that OLFM4 is associated with cathepsin C in neutrophils and suppresses its activity, we investigated whether the enhanced neutrophil bacteria-killing capability in *OLFM4*^{-/-} mice was accompanied by enhanced cathepsin C activity. The cathepsin C activity in neutrophils from

OLFM4^{-/-} mice was significantly increased relative to that of neutrophils from WT mice under both noninfection and infection conditions (Fig. 4). The activities of the serine proteases neutrophil elastase (Fig. 5A), proteinase 3 (Fig. 5B), and cathepsin G (Fig. 5C) were also significantly higher in neutrophils from *OLFM4*^{-/-} mice compared with those isolated from WT mice. These results suggest that loss of OLFM4 in neutrophils leads to enhanced cathepsin C and serine protease activities, thereby increasing bacterial killing.

Cathepsin C contributes to enhanced bacterial killing in *OLFM4*^{-/-} mice

To confirm that cathepsin C plays an essential role in OLFM4's downregulation of neutrophil bacterial killing, we created *OLFM4*^{-/-} *CatC*^{-/-} mice and cathepsin C double-deficient mice (*OLFM4*^{-/-} *CatC*^{-/-}). The intracellular bacterial killing activity observed in *OLFM4*^{-/-} mice that was enhanced relative to WT mice was significantly com-

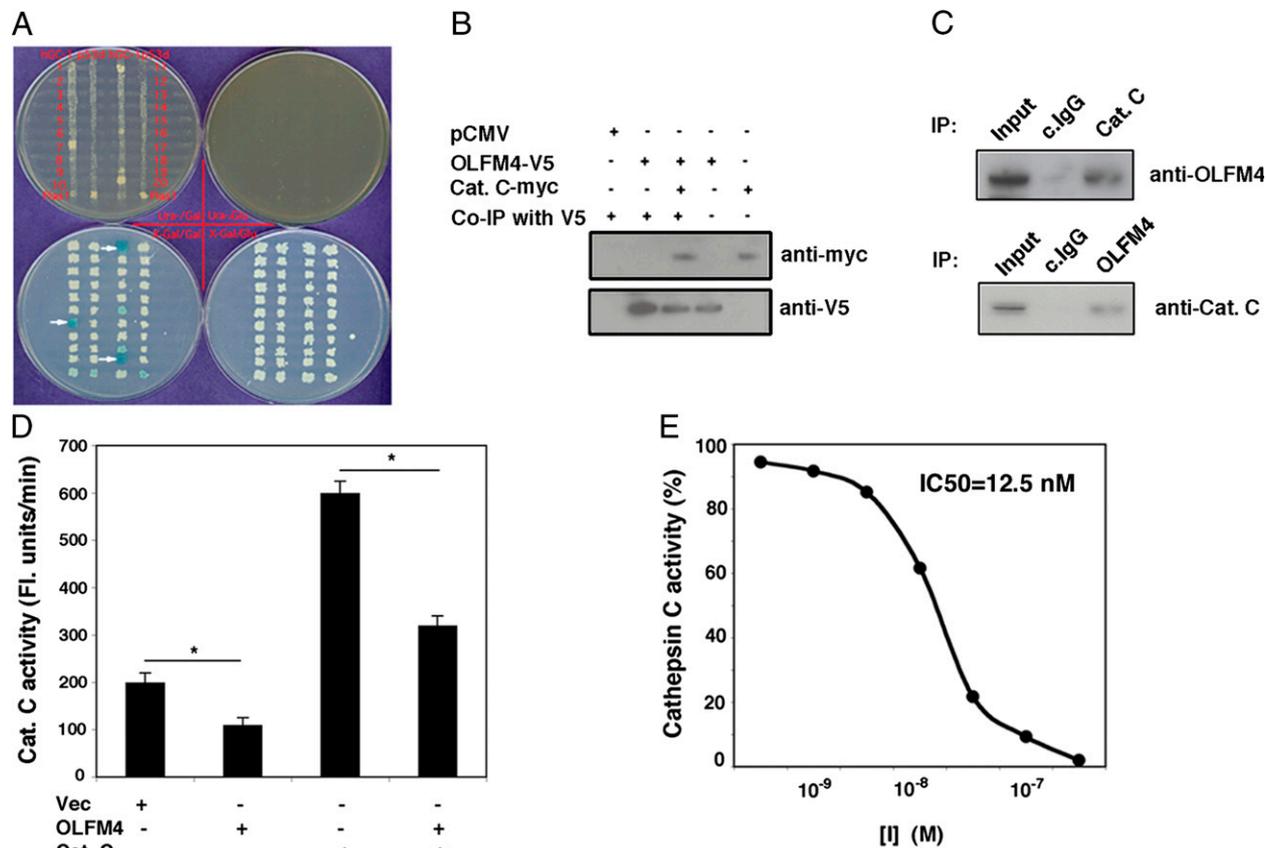


FIGURE 3. OLFM4 binds cathepsin C and is an inhibitor of cathepsin C in vitro and in vivo. **(A)** A yeast two-hybrid assay was performed to identify potential binding partners of OLFM4. The blue colonies (indicated by white arrows) were isolated and confirmed to be cathepsin C by sequencing. **(B)** Expression plasmids of OLFM4-V5 and cathepsin C-myc were transfected individually or cotransfected into HEK 293T cells. After 24 h, the total cell lysates were immunoprecipitated with V5 Ab and probed with myc or V5 Ab by Western blotting. **(C)** Total cell lysates from normal human bone marrow neutrophils were immunoprecipitated with cathepsin C Ab (Cat. C) or control IgG Ab (c.IgG) and probed with OLFM4 Ab by Western blotting (*top panel*). Total cell lysates from normal human bone marrow neutrophils were immunoprecipitated with OLFM4 Ab or c.IgG and probed with Cat. C Ab by Western blotting (*bottom panel*). Lysate before immunoprecipitation was loaded as a control (Input). **(D)** HEK 293T cells were transfected with OLFM4-V5, cathepsin C-myc, or control vector (Vec) expression plasmid, or cotransfected with OLFM4-V5 and cathepsin C-myc expression plasmids. Cathepsin C activity was analyzed by hydrolyzing Gly-Arg-AMC. Results are expressed as mean \pm SD ($n = 4$) fluorescence units per minute. * $p < 0.05$. **(E)** Cathepsin C (250 ng/ml) was incubated with purified OLFM4 protein at various concentrations as indicated at room temperature for 30 min, and the cathepsin C activity was analyzed. The percentage of cathepsin C activity was demonstrated.

promised by the additional loss of cathepsin C in mice with OLFM4 and cathepsin C double deficiency ($OLFM4^{-/-}CatC^{-/-}$; Fig. 6A). Similarly, double deficiency of OLFM4 and cathepsin C also significantly reduced the in vivo bacterial clearance observed in $OLFM4^{-/-}$ mice that was enhanced relative to WT mice (Fig. 6B). In agreement with a previous report that cathepsin C-deficient mice have impaired bacterial clearance (20), the intracellular bacteria killing and clearance of *S. aureus* and *E. coli* were found to be significantly impaired (Fig. 6A, 6B). That the additional loss of cathepsin C in $OLFM4^{-/-}$ mice ($OLFM4^{-/-}CatC^{-/-}$) led to a compromised bacterial clearance, but not to the extent of defect in $CatC^{-/-}$ mice (Fig. 6A, 6B), suggested that OLFM4 modulates bacterial clearance through both a cathepsin C-dependent and -independent mechanism. These data provide direct evidence that there is a functional interplay between OLFM4 and cathepsin C gene, and OLFM4 restriction of neutrophil bacterial killing is mediated, at least partially, through interaction with cathepsin C.

Discussion

S. aureus and *E. coli* infections are still major threats to human health, especially because of emerging antimicrobial resistance (21). Neutrophils play essential roles in killing invading bacteria. Therefore, it is important to understand the molecular mechanisms

underlying neutrophil activity to better combat bacteria infection. In this study, we demonstrate that OLFM4 is a critical regulator of neutrophil's ability to kill *S. aureus* and *E. coli*. This function is associated with restriction of cathepsin C and downstream serine proteases, but not with NADPH-mediated oxidative burst.

We demonstrate that OLFM4 physically associates with cathepsin C in neutrophils. We show that OLFM4 is localized in multiple granule compartments with most abundance in specific granules, but also in azurophilic granules where cathepsin C and serine proteases are mostly localized (15). As previously described, there is a functional interplay between different granule subsets (22). For example, neutrophil gelatinase-associated lipocalin that is normally found in the specific granules is able to complex with gelatinase (23). Proteases from azurophilic granules may activate the cathelicidin present in specific granules (22). Therefore, OLFM4 abundantly present in specific granules may also interact with cathepsin C in azurophilic granules, especially when azurophilic granules and specific granules are fused with phagosomes to form phagolysosomes after bacteria phagocytosis by neutrophils.

Recent studies showed that cathepsin C is involved in multiple pathological processes, particularly in neutrophils where it plays an important role in bacterial clearance (20) and the inflammatory response (24). Thus, cathepsin C represents a potential therapeutic

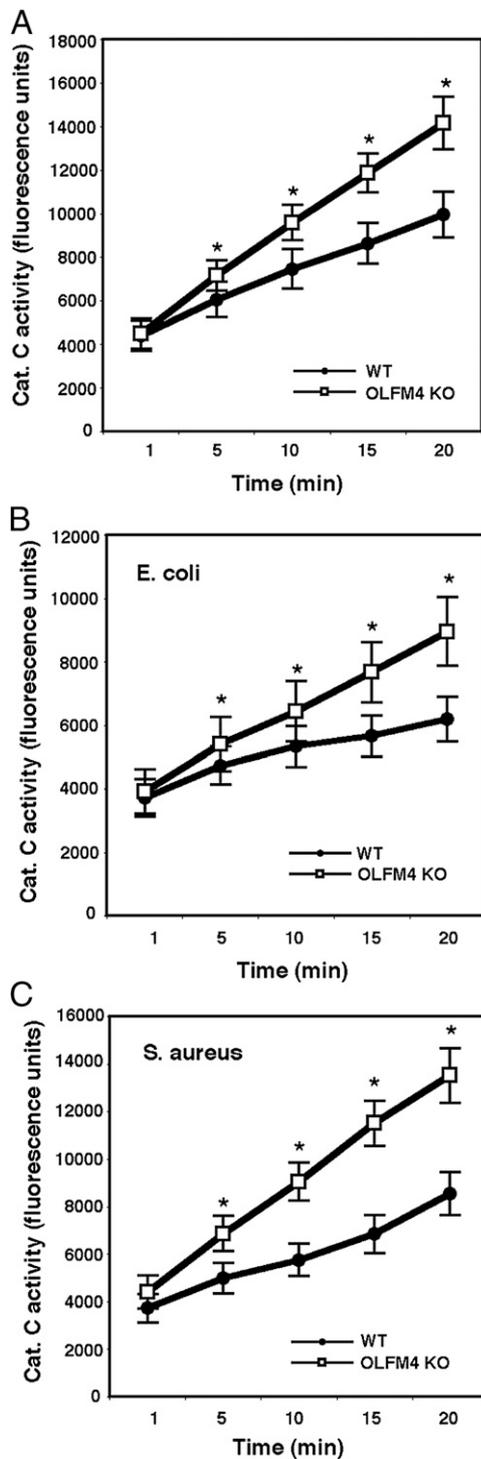


FIGURE 4. Neutrophils from *OLFM4*^{-/-} mice display enhanced activities of cathepsin C. Neutrophils (5×10^6) derived from bone marrow of WT and *OLFM4*^{-/-} (*OLFM4* KO) mice were incubated in vitro without (A) or with *E. coli* (5×10^7) (B) or *S. aureus* (5×10^7) (C) for 1 h. Cells were lysed in cathepsin C lysis buffer, and equal amounts of lysates were used for cathepsin C activity assays. The cathepsin C activities in neutrophils from *OLFM4* KO mice were significantly higher than those from WT mice ($*p < 0.01$) in the absence or presence of bacterial challenge. Data are expressed as mean \pm SD ($n = 3$).

target for the treatment of inflammatory disease. Many small peptides have been selected to be potent cathepsin C inhibitors, but they have poor stability and/or pharmacokinetics (25). Cathepsin C is also inhibited by protein inhibitors, such as rat stefin

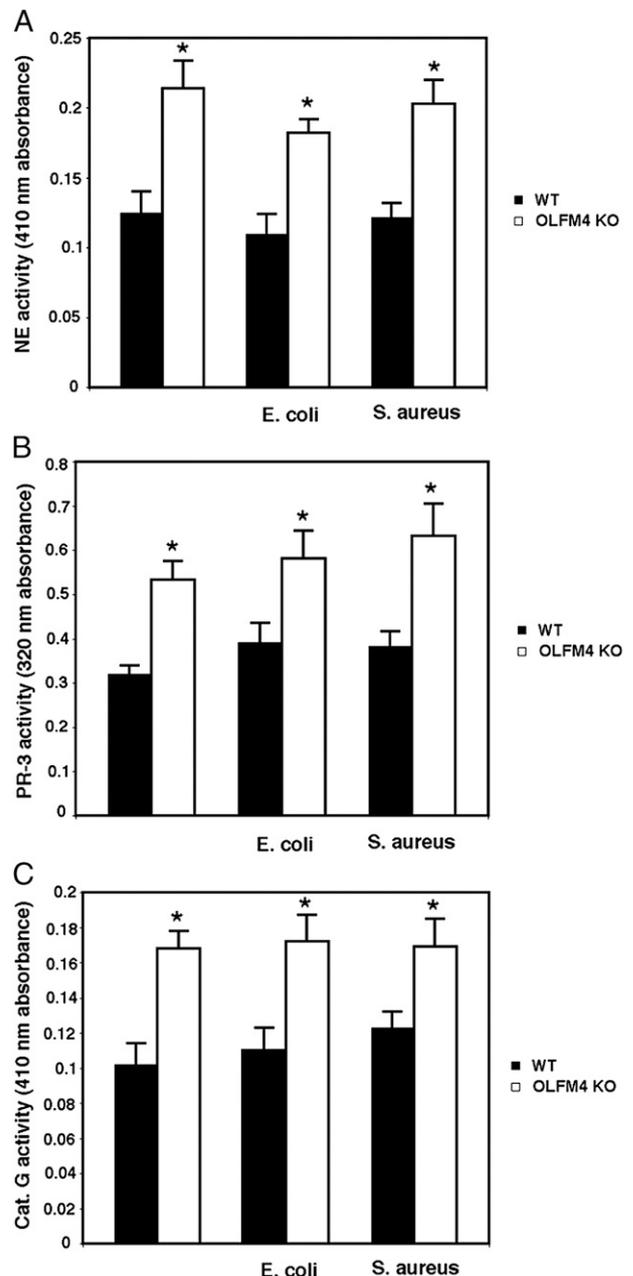
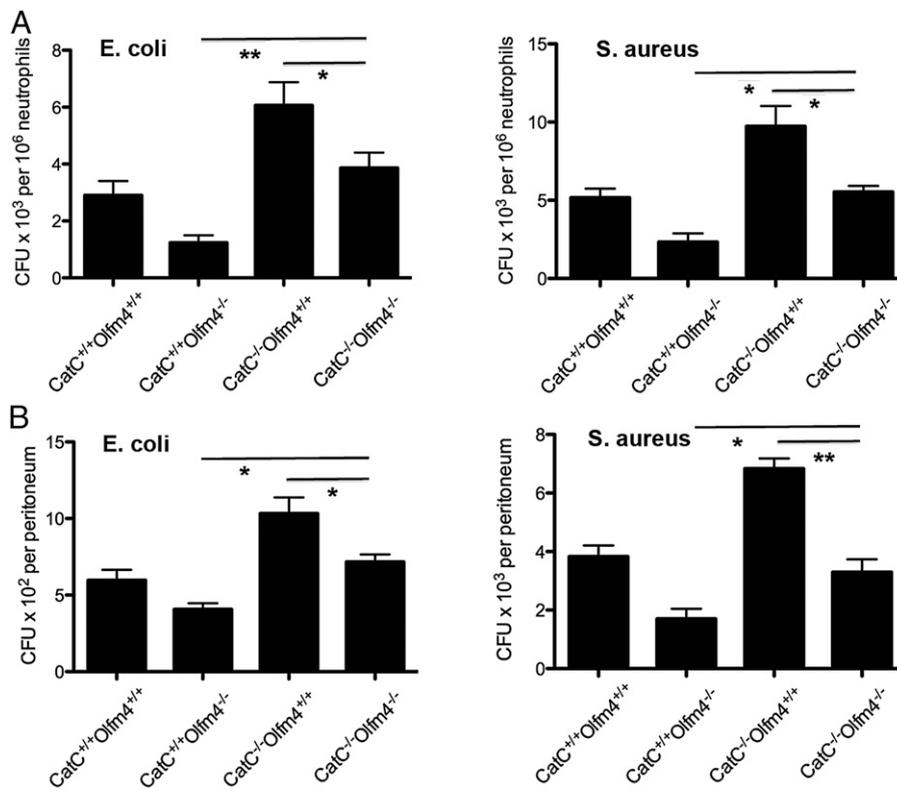


FIGURE 5. Neutrophils from *OLFM4*^{-/-} mice display enhanced activities of serine proteases. (A) Cells were lysed in serine protease lysis buffer, and equal amounts of lysates were used for neutrophil elastase (NE) activity assay using methoxysuccinyl-Ala-Ala-Pro-Val-*p*-nitroanilide as a substrate at room temperature for 30 min. The absorbance at 410 nm was measured. $*p < 0.05$ when compared with WT. (B) Cells were lysed in serine protease lysis buffer, and equal amounts of lysates were used for proteinase-3 (PR-3) activity assay using Boc-Ala-Ala-Nva-SBzL as a substrate at room temperature for 30 min. The absorbance at 320 nm was measured. $*p < 0.05$ when compared with WT. (C) Cells were lysed in serine protease lysis buffer, and equal amounts of lysates were used for cathepsin G (Cat. G) activity assay using *N*-succinyl-Ala-Ala-Pro-Phe-*p*-nitroanilide as a substrate at room temperature for 30 min. The absorbance at 410 nm was measured. $*p < 0.05$ when compared with WT. All data are expressed as mean \pm SD ($n = 3$).

A and chicken cystatin F, two important inhibitors of cysteine peptidases from the cystatin superfamily (26, 27). Cystatin F is expressed primarily in CD8⁺ T cells and in CD56⁺ NK cells, and has been shown to complex with cathepsin C in human monocytic and NK cells (27). Our study showed that *OLFM4*, a granule

FIGURE 6. Enhanced in vitro intracellular bacterial killing and in vivo clearance observed in *OLFM4*^{-/-} mice are compromised in cathepsin C and OLFM4 double-deficient mice. **(A)** Bone-marrow neutrophils (10⁶) from different genotype mice as indicated were incubated with preopsonized *E. coli* (10⁷) or *S. aureus* (10⁷) in a 24-well plate. After 15 min, the media were gently removed. The cells were treated with gentamicin (100 μg/ml) for *E. coli* or lysostaphin (50 μg/ml) for *S. aureus* for an additional 30 min. The viable bacteria were counted on tryptic soy agar plates. Data are expressed as mean ± SD (*n* = 3). **p* < 0.05, ***p* < 0.01. **(B)** Mice were challenged i.p. with *E. coli* (5 × 10³) or *S. aureus* (5 × 10⁴). After 2 h, the peritoneal cavity was lavaged and the number of viable bacteria was determined using the standard plate method. Data are expressed as mean ± SD (*n* = 3). **p* < 0.05, ***p* < 0.01.



protein, was associated with cathepsin C in neutrophils, and strongly and specifically inhibited cathepsin C both in vitro and in vivo. Moreover, the data from the *OLFM4*^{-/-} mouse model demonstrated that OLFM4 also affects multiple serine protease activities indirectly. Thus, OLFM4 represents a novel protein inhibitor of cathepsin C, specifically in neutrophils.

Our results show that enhanced bacteria killing activity in OLFM4-deficient neutrophils is associated with increased cathepsin C and downstream serine protease activities. It has been reported that bacterial clearance is impaired in cathepsin C^{-/-} mice using a cecal ligation and puncture model of septic peritonitis (20), demonstrating that cathepsin C is an important regulator for bacteria control and clearance. Although a regulatory role of cathepsin C on specific bacteria such as *S. aureus* and *E. coli* used in this study has not been reported, mice clearance of *E. coli* and *S. aureus* are impaired in absence of active neutrophil elastase and cathepsin G, which requires activation from cathepsin C (1). The mode of action of OLFM4 through cathepsin C is further confirmed by experiments in OLFM4 and cathepsin C double-deficient mice. The enhanced bactericidal abilities in the neutrophils of *OLFM4*^{-/-} mice were compromised, but not eliminated, by further deletion of cathepsin C gene, confirming the involvement of cathepsin C in the OLFM4-mediated effect. This suggests that the increased bactericidal activity in *OLFM4*^{-/-} neutrophils is, at least in part, due to the enhanced cathepsin C activity. However, these data do not rule out possible involvement of other mechanisms in the observed phenotypes. For example, it was previously reported that OLFM4 negatively regulates the nucleotide-binding oligomerization domain (Nod) 1 and Nod2 (12). Nod1 has a broad influence on neutrophil function, activating both oxidative and nonoxidative mechanisms of killing (28). Therefore, the increased bactericidal activity in OLFM4-deficient neutrophils may also be partially caused by subsequent enhanced Nod1 activity in neutrophils.

OLFM4 and other olfactomedin-related proteins have a high m.w. multimer structure with intramolecular and intermolecular disulfide bonds (13). It has been previously reported that OLFM4 complexes with cadherin (13) and Nods (12) proteins. These observations together with this study suggest a possibility that OLFM4 may be a component of multiple protein complexes as an adaptor or scaffold protein. The notion that olfactomedin class of proteins may be scaffolds for proteases and their substrates was also recently suggested by Inomata and coworkers (29) based on their finding that a *Xenopus* secreted olfactomedin 1 recruits the Tollid proteases to their substrate chordin and is required for normal chordin degradation.

In summary, we demonstrate that OLFM4 is a neutrophil granule protein, which proved to be an important negative regulator of bacteria killing as evidenced by that neutrophils from *OLFM4*^{-/-} mice have increased capability to kill *S. aureus* and *E. coli*, and *OLFM4*^{-/-} mice are more resistant to systemic sepsis. Taken together with the previous study (12), which demonstrated that OLFM4 downregulates host innate immunity against *H. pylori*, OLFM4 appears to be an important regulator of host innate immunity against a broad array of bacterial infections. OLFM4 may prove to be a potential target for therapeutic augmentation of host innate immunity in genetic immune-deficient patients such as those with chronic granulomatous disease.

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Disclosures

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