



## Multiplex PCR Master

Master mix for multiplex PCR

Cat. No.	Amount
PCR-110S	2 x 1,25 ml (2x conc.)
PCR-110L	10 x 1,25 ml (2x conc.)

**For general laboratory use.**

**Shipping:** shipped on gel packs

**Storage Conditions:** store at -20 °C

**Additional Storage Conditions:** avoid freeze/thaw cycles

**Shelf Life:** 12 months

**Form:** liquid

**Concentration:** 2x conc.

### Description:

Multiplex PCR Master is specially designed for the set-up of multiplex PCR reactions. It contains an optimized composition of polymerase, nucleotides, MgCl<sub>2</sub> and stabilizing components in a specifically developed buffer system allowing the parallel amplification of a multitude of fragments in a single PCR assay.

The master mix contains all reagents (except primer and template) in a 2x concentrated ready-to-use solution.

The kit is recommended for use in routine PCR reactions and highly suitable for multiple target gene amplification in a single tube.

The high specificity and sensitivity of the mix is achieved by a chemically inhibited hot-start polymerase. Its activity is blocked at ambient temperature preventing the extension of nonspecifically annealed primers and primer-dimer formations at low temperatures during PCR setup.

### Content:

#### 2x Multiplex PCR Master (red cap)

master mix containing Hot Start Taq polymerase, nucleotides, optimized reaction buffer and stabilizers

#### PCR grade water (white cap)

### Recommended 50 µl PCR assay:

Prepare a master mix of all components except template to reduce pipetting errors. A reaction volume of 20-50 µl per assay is recommended for most PCR cyclers. Pipet with sterile filter tips and perform the setup in an area separate from DNA preparation or analysis. No-template controls should be included in all amplifications.



## Multiplex PCR Master

Master mix for multiplex PCR

component	stock conc.	final conc.	50 µl assay
Multiplex PCR Master	2x	1x	25 µl
forward primer 1	10 µM	400 nM	2 µl
reverse primer 1	10 µM	400 nM	2 µl
forward primer 2	10 µM	400 nM	2 µl
reverse primer 2	10 µM	400 nM	2 µl
forward primer ...	10 µM	400 nM	2 µl
reverse primer ...	10 µM	400 nM	2 µl
Template a) animal genomic DNA b) bacterial genomic DNA c) plasmid and lambda DNA	-	-	a) 10-200 ng b) 1 - 50 ng c) 1 - 5 ng
PCR-grade water	-	-	fill up to 50 µl

<sup>2)</sup>Cycle numbers are recommended as following:

- animal genomic DNA  
10 - 50 ng: 35 - 50 cycles  
50 - 200 ng: 30 - 45 cycles
- bacterial genomic DNA  
1 - 5 ng: 35 - 50 cycles  
5 - 50 ng: 30 - 40 cycles
- plasmid and lambda DNA  
1 - 5 ng: 30 - 40 cycles

<sup>3)</sup>The elongation time depends on the length of the fragments to be amplified. A time of 1 min/kb is recommended.

For optimal specificity and amplification an individual optimization of the recommended parameters may be necessary for each new template DNA and/or primer pair.

### Recommended cycling conditions:

Initial denaturation	95 °C	12 min	1x
Denaturation	95 °C	30 sec	30 - 50x <sup>2)</sup>
Annealing <sup>1)</sup>	58 - 64 °C	40 sec	30 - 50x <sup>2)</sup>
Elongation <sup>3)</sup>	72 °C	1 min/kb	30 - 50x <sup>2)</sup>
Final elongation	72 °C	5 min	1x

<sup>1)</sup>The optimal annealing temperature (AT) can be calculated for each primer as following:

$$AT = T_m - 5\text{ °C with } T_m = 2\text{ °C} \times (A+T) + 4\text{ °C} \times (G+C)$$

Please note that primers should be designed to show minimal differences in their melting temperatures ( $T_m$ ).