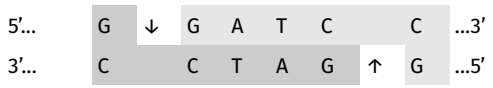
DATA SHEET

Jena Bioscience

BamHI

JBSpeed Restriction Enzyme

Cat. No.	Amount
EN-103S	7.500 Units
EN-103L	5 x 7500 Units



Unit Definition: One unit is the amount of enzyme required to completely digest 1 µg of Lambda DNA (5 sites) in 1 hour in a total reaction volume of 50 µl. Enzyme activity was determined in the recommended reaction buffer.

For general laboratory use.

Please centrifuge briefly before opening (volume ≤2 ml).

Shipping: shipped on gel packs

Storage Conditions: store at -20 °C

Additional Storage Conditions: avoid freeze/thaw cycles

Shelf Life: 12 months

Form: liquid

Concentration: 10 units/µl

Source: *Bacillus amyloliquefaciens* H

Supplied with: 10x Buffer BamHI

Recommended 50 µl assay

5 µl	10x Buffer BamHI
1 µg	pure DNA[1] or PCR product[2]
10 units	enzyme
fill up to 50 µl	PCR grade water

[1] Supercoiled or high molecular weight DNA (e.g. plant genomic DNA) may require longer incubation time or higher amount of enzyme.
[2] Some enzymes may require additional DNA bases flanking the restriction site for complete digestion.

Protocol:

- The enzyme should not exceed 10 % of total reaction volume.
- Add enzyme as last component. Mix components well before adding enzyme. After enzyme addition, mix gently by pipetting. Do not vortex.
- Incubate 5 to 10 min. at 37 °C.
- Stop reaction by alternatively:
  - Addition of 2.1 µl EDTA pH 8.0 [0.5 M], final 20 mM
  - Heat Inactivation (20 min. at 80 °C)
  - Spin Column DNA Purification (e.g. PCR Purification Kit, Cat.-No. PP-201S/L)
  - Gel Electrophoresis and Single Band Excision (e.g. Agarose Gel Extraction Kit, Cat.-No. PP-202S/L)
  - Phenol-Chloroform Extraction or Ethanol Precipitation.

Please note that the optimum digestion condition for this enzyme is 1x Buffer BamHI. Within the Universal Buffer (UB) system, the most majority of our enzymes display 100% Relative Activity in 1x UB and only few either in 0.5x or 2x UB. If optimum condition for second enzyme is different than the recommended for the first enzyme, we suggest carrying out first the restriction at the higher recommended concentration of UB and dilute the reaction volume to the adequate UB concentration for further proceeding with the second restriction.

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Reaction Enzymes Buffer Guide:

Buffer 1	10 × B1	100 mM 100 mM 1000 µg/ml	Tris-HCl (pH 7.9, 25°C) $MgCl_2$ BSA
Buffer 2	10 × B2	100 mM 100 mM 500 mM 1000 µg/ml	Tris-HCl (pH 7.9, 25°C) $MgCl_2$ NaCl BSA
Buffer 3	10 × B3	500 mM 100 mM 1000 mM 1000 µg/ml	Tris-HCl (pH 7.9, 25°C) $MgCl_2$ NaCl BSA
Buffer 4	10 × B4	100 mM 100 mM 1500 mM 1000 µg/ml	Tris-HCl (pH 7.9, 25°C) $MgCl_2$ NaCl BSA
Buffer 5	10 × B5	200 mM 100 mM 500 mM 1000 µg/ml	Tris-acetate (pH 7.9, 25°C) Mg-acetate K-acetate BSA

Reaction Buffer Compatibility:
Our restriction enzymes are fully compatible to restrictases and buffer systems from other manufacturers and can be used along in double digestions. To obtain best results, consult the corresponding manuals of all involved products.

Ligation and recutting:
After 50-fold overdigestion with BamHI, >95 % of the DNA fragments can be ligated and recut with this enzyme.

Star activity:
Conditions of low ionic strength, high enzyme concentration, glycerol concentration >5 % or pH >8.0 may result in star activity.

DNA Methylation:
No Inhibition: dam, dcm, CpG

Quality Control:
All preparations are assayed for contaminating endonuclease, 3'-exonuclease, 5' exonuclease/ 5' phosphatase, as well as nonspecific single- and doublestranded DNase activities.

Related Products:
Universal Restriction Buffer, #EN-300