

Taq Master Lyophilisate

Lyophilized Taq master mix

Ready-to-use lyophilisates for PCR

Size	Cat.-No.	Amount
8-tube strips	PCR-152S-8TS	12 strips / 96 reactions
	PCR-152L-8TS	60 strips / 480 reactions
96-well plates (flat top, without skirt)	PCR-152S-FTP	2 plates / 192 reactions
	PCR-152L-FTP	10 plates / 960 reactions
96-well plates (half skirt)	PCR-152S-HSP	2 plates / 192 reactions
	PCR-152L-HSP	10 plates / 960 reactions

For *in vitro* use only

Quality guaranteed for 6 months when stored below 20°C

Taq Master Lyophilisate

Preloaded lyophilisates of Taq DNA polymerase, dATP, dCTP, dGTP, dTTP, reaction buffer with MgCl₂, and stabilizers

PCR-grade water

Description

Taq Master Lyophilisate is delivered in PCR reaction tube strips or 96-well plates preloaded with a complete master mix in a dry, room temperature stable format. The lyophilisate combines highest performance with convenience of use and stability. There is no need for freezing, thawing or pipetting on ice. The few remaining pipetting steps minimize the risk of errors or contaminations.

Each vial contains polymerase, dNTPs and reaction buffer with MgCl₂ required for a 20 µl PCR assay. To perform PCR, fill up the vials with a premix of primers and PCR-grade water and add DNA template. If necessary, centrifuge to remove bubbles, vortex the vials to assure homogeneity and start cycling.

Recommended PCR assay

20 µl PCR assay		
forward Primer	0.2-1 µM	0.4-2 µl / 10 µM
reverse Primer	0.2-1 µM	0.4-2 µl / 10 µM
Template DNA	1-50 ng	
PCR grade H ₂ O	fill up to 20 µl	

Recommended cycling conditions

Initial denaturation	94°C	2 min	1x
Denaturation	94°C	30 sec	30x
Annealing ¹⁾	45 - 68°C	30 sec	
Elongation ²⁾	72°C	30 sec - 3 min	
Final elongation	72°C	2 min	1x

1) The annealing temperature depends on the melting temperature of the primers used.

2) The elongation time depends on the length of the fragments to be amplified. A time of 1 min/kbp is recommended.

For optimal specificity and amplification an individual optimization of the recommended parameters may be necessary for each new primer-template pair.