



LEXSY

the protein expression platform based on

Leishmania tarentolae

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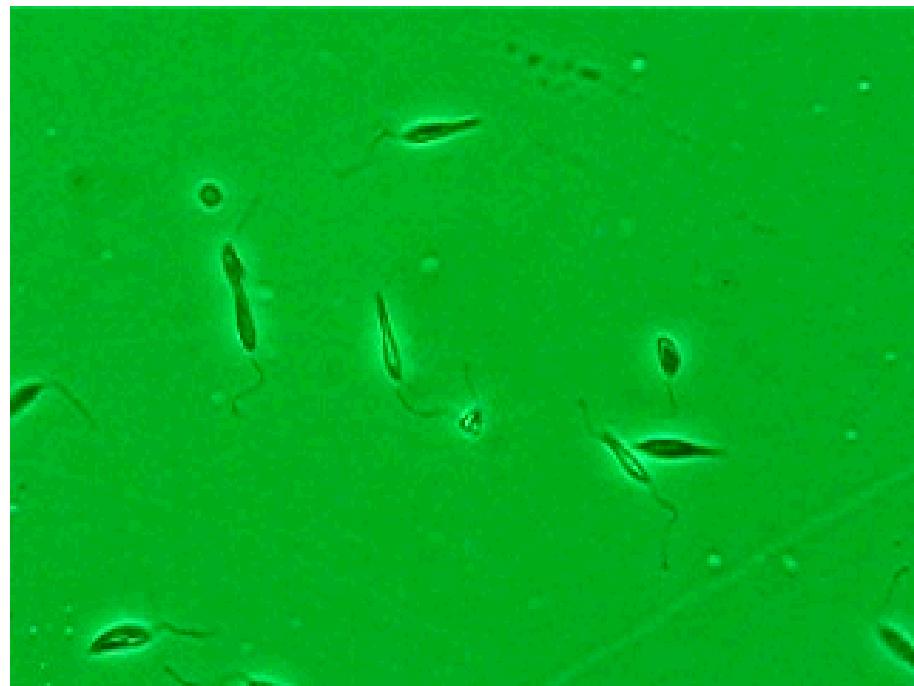
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Leishmania tarentolae was chosen as expression host for production of recombinant proteins



Host: *Tarentola mauritanica*

- unicellular flagellated protozoa
- parasite of lizard (*Sauroleishmania*)
- not pathogenic for mammals
- **biosafety group 1 organism**
- can be easily cultured *in vitro*
- genome sequence published 2012
- transcriptomic and proteomic data 2015

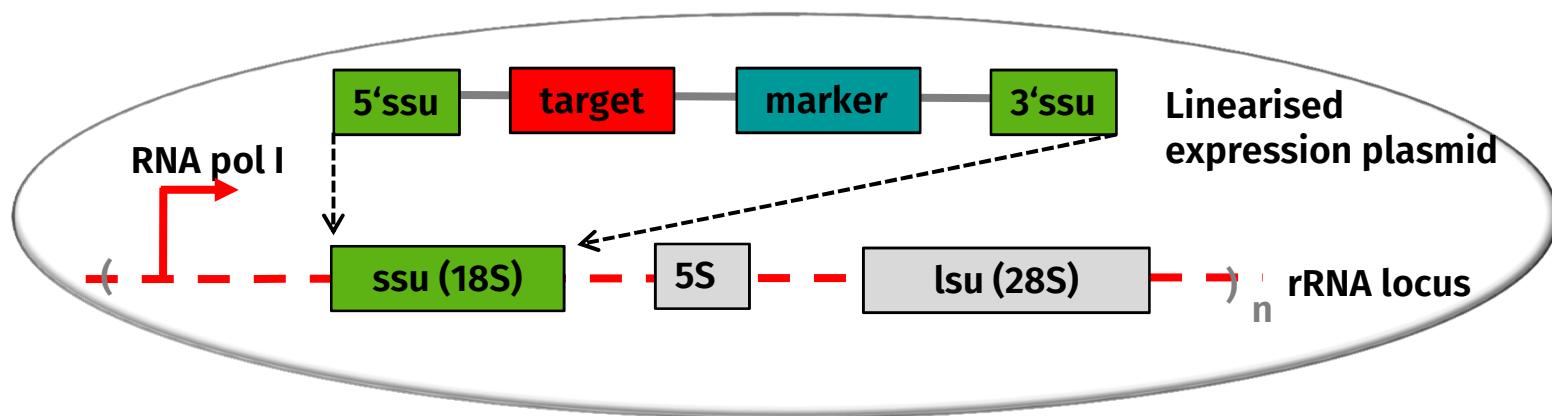


“LEXSY”

Leishmania
Expression
System

Configuration 1: Constitutive expression

- **ssu integration (multicopy 18S rRNA cluster)**
- **transcription by host RNA polymerase I**

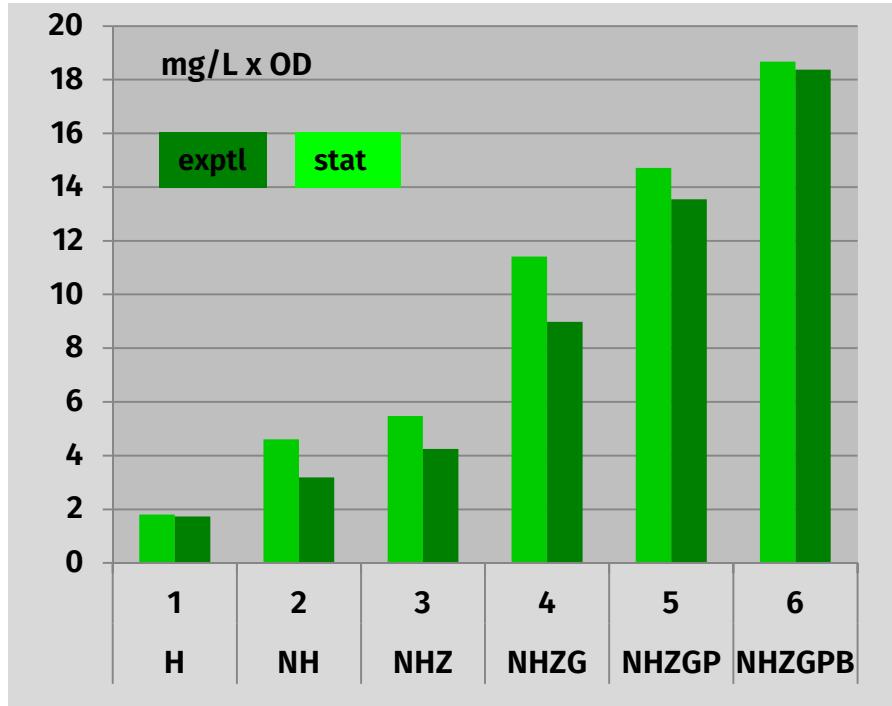


- **RNA polymerase I: 10x stronger than RNA Polymerase II**
- **maximal activity during exponential growth (early product synthesis)**
- **six selection markers (Nourseothricin, Blasticidin, Bleomycin, Genitycin G418, Hygromycin & Puromycin)**

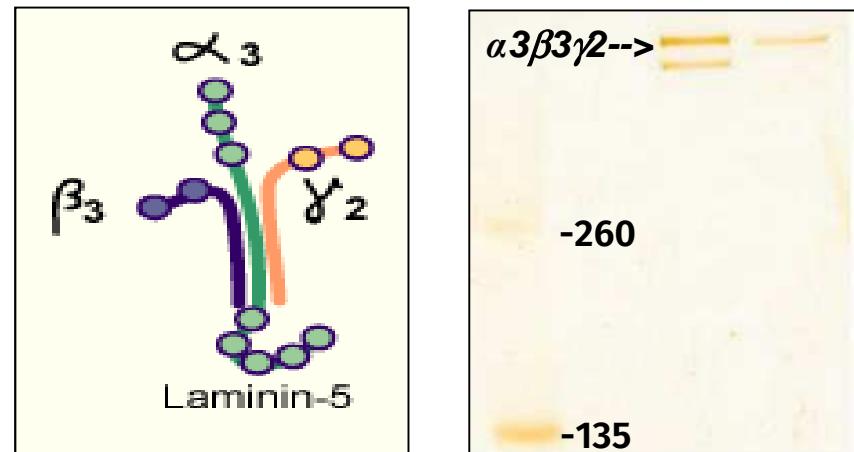
Worked also in *L. amazonensis*, *L. donovani*, *L. infantum*, *L. major*, *L. mexicana*,
Phytomonas serpens ...

Co-integration of expression cassettes into *ssu* locus

Additive effect on protein expression



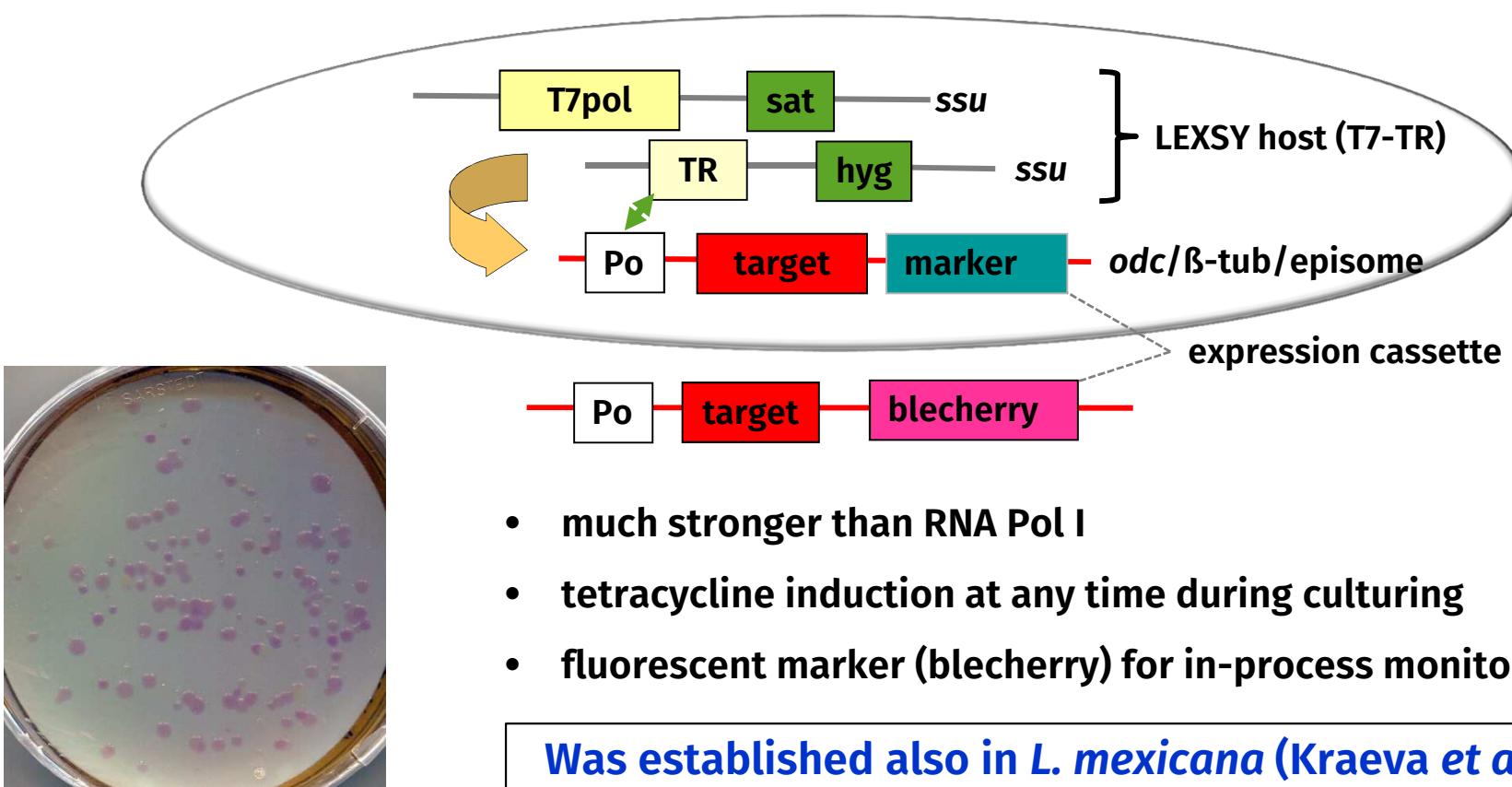
Expression of multi-subunit proteins



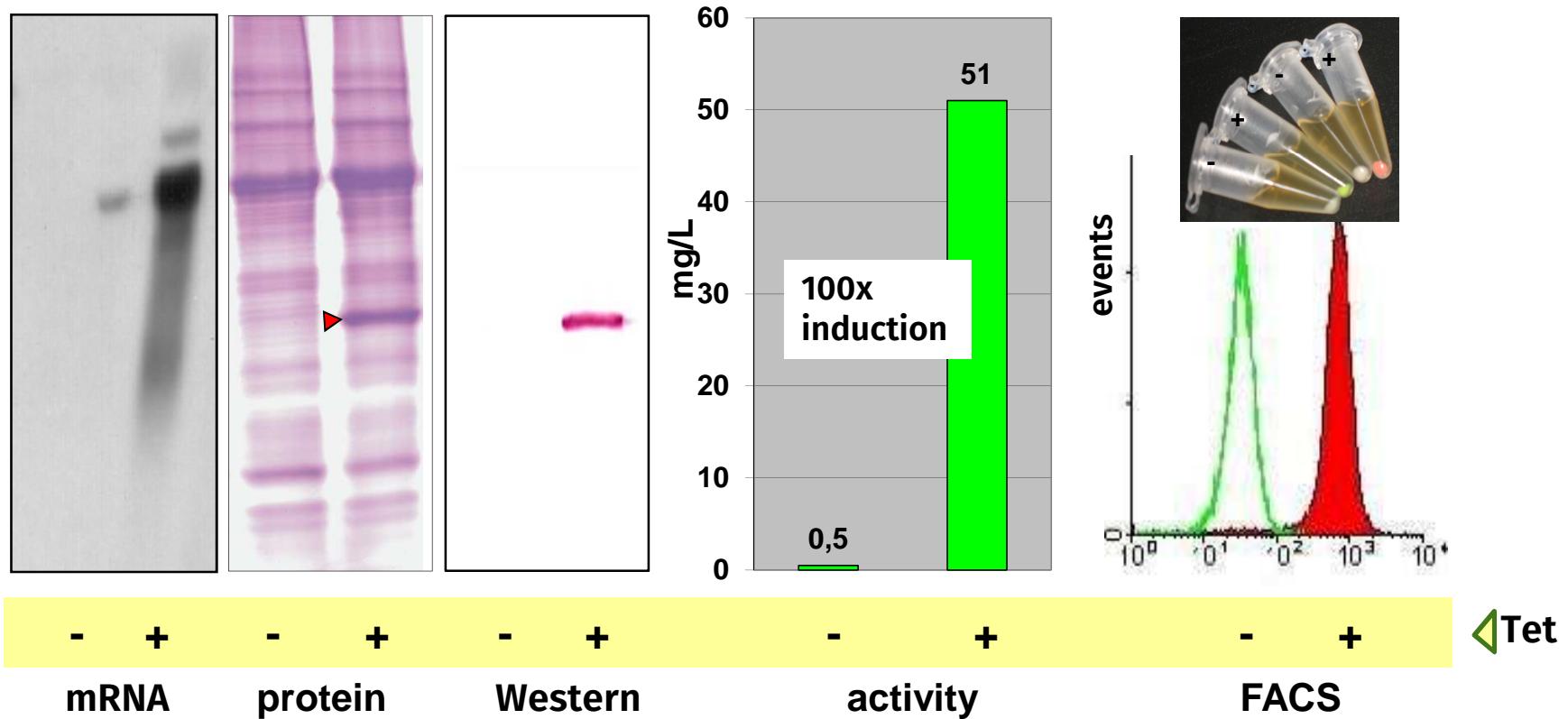
420 kDa Laminin-332 heterotrimer
secreted from LEXSY
Phan *et al.* (2009) *Protein Expression and Purification* 68:79.

Configuration 2: Inducible expression

- transcription by heterologous T7 RNA polymerase
- control by TET repressor
- *odc* (β -tub) integration or episomal propagation of target gene

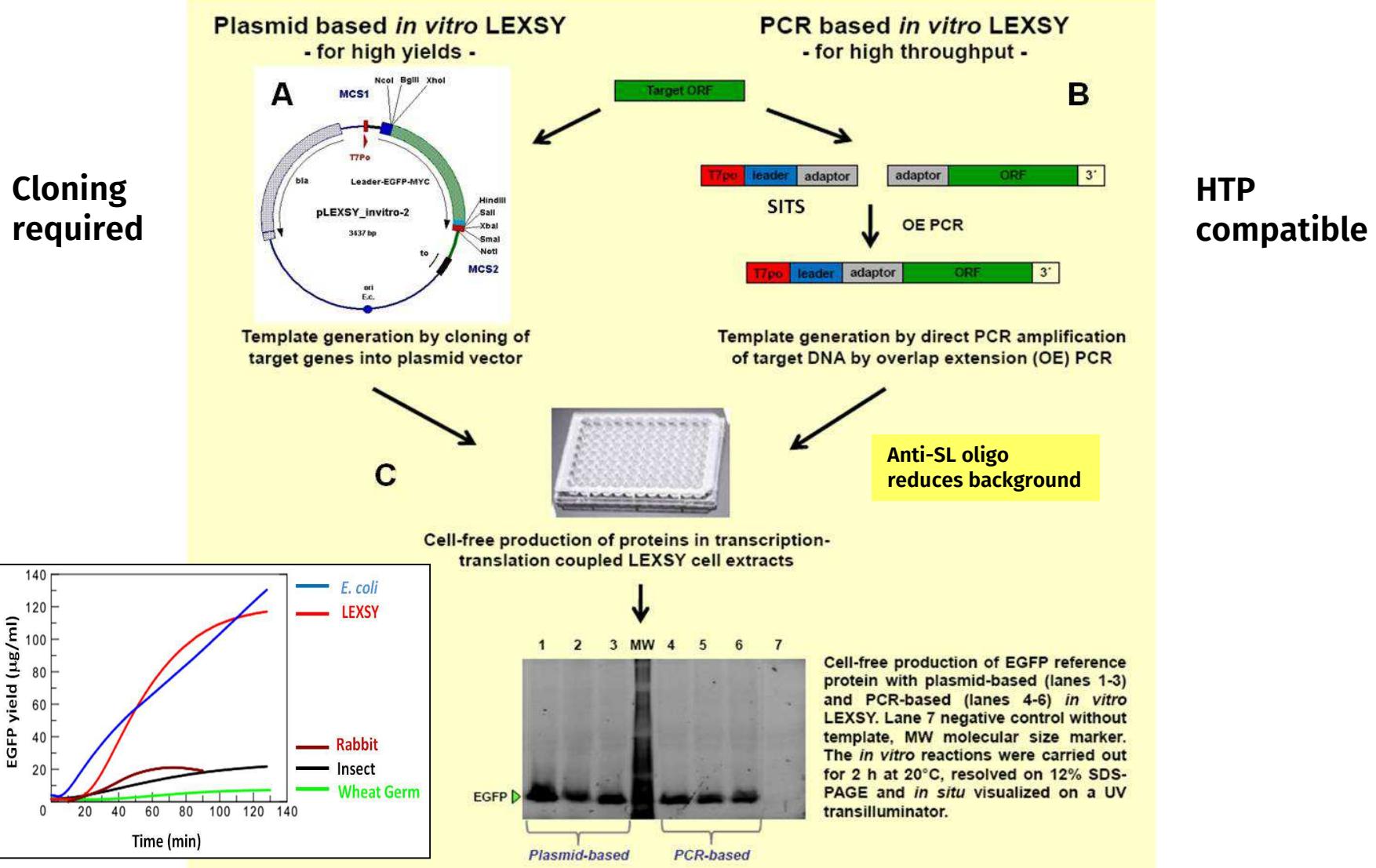


Efficient expression control in inducible LEXSY from mRNA to protein



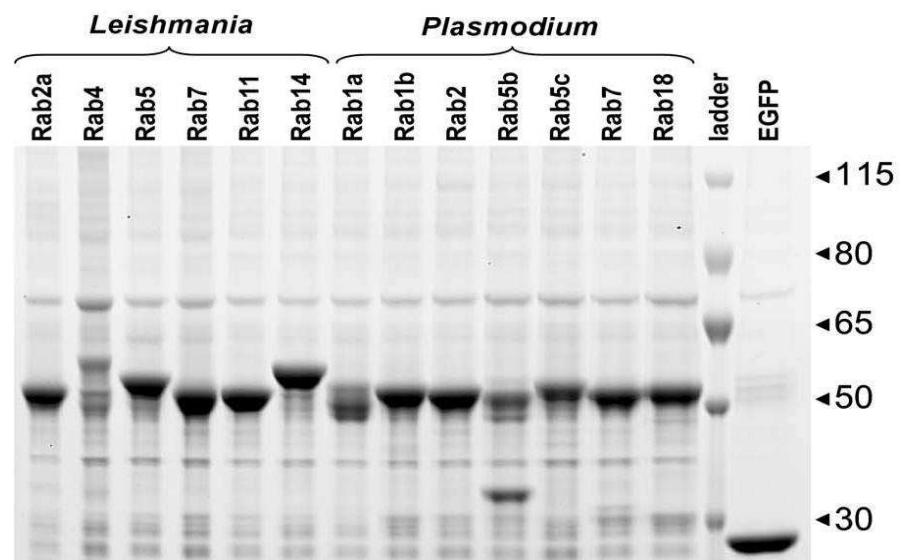
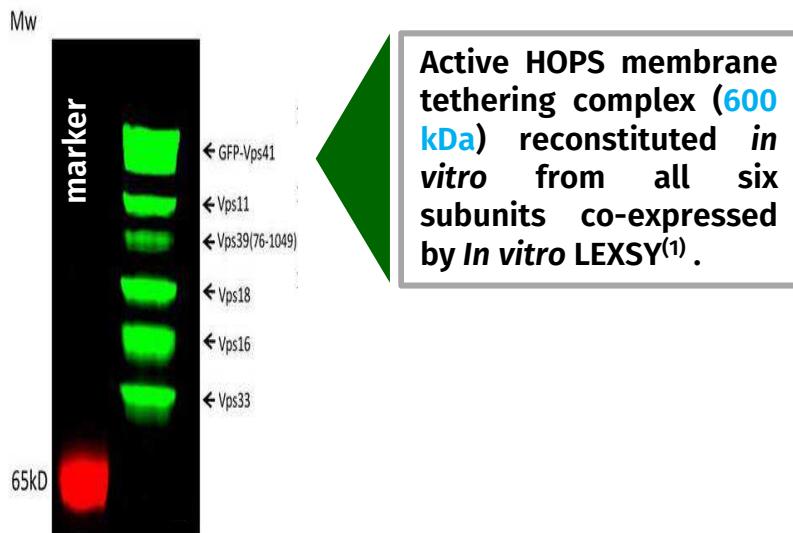
- induction profiles and yields stable over > 500 generations
- all cells induced

In Vitro LEXSY: Rapid cell-free protein production



Versatility of In Vitro LEXSY

- LEXSY cell-free production of to date largest multisubunit membrane protein complex⁽¹⁾
- Fluorescence-based protein interaction studies without purification^(2,3,4)
- Fast high throughput screening^(5,6,7)



- (1) Guo *et al.* (2013) PLoS ONE 8: e8153
(2) Brooks *et al.* (2014) Science 344: 1249783
(3) Gambin *et al.* (2014) eLife 3: e01434
(4) Han *et al.* (2014) Journal of Biological Chemistry 289: 7764
(5) Mureev *et al.* (2009) Nature Biotechnology 27: 747
(6) Kovtun *et al.* (2010) PLOS one 5: e14388
(7) Kovtun *et al.* (2011) Methods 55: 58

RabGTPases from *L. tarentolae* or *P. falciparum* were expressed in PCR-based *In Vitro* LEXSY. Coomassie stained SDS-PAGE gel loaded with EGFP-Rab GTPases eluted from GFP-Cap matrix (adapted from Kovtun *et al.* 2010).

LEXSY – work horse for parasitologists due to phylogenetic relationship to other Kinetoplastids

Expression of kinetoplastid proteins in *L. tarentolae* e.g.

- Surface antigens of *T. brucei gambiense* ISG65, VSG LiTat 1.3 and VSG LiTat 1.5 (Rooney et al. 2015)
- Surface antigen PSA-38S of *L. amazonensis* (Chamakh-Ayari et al. 2014)
- α -N-acetylglucosaminyltransferase OGNT2 from *T. cruzi* (Heise et al. 2009)

Genetic manipulation of *Leishmania* species e.g.

- Fluorescent/luminiscent cells (Lang et al. 2005, Barak et al. 2005..... Sadlova et al. 2011, Bolhassani et al. 2011 , Rabhi et al. 2015, Taheri et al. 2015, García-Hernández et al. 2015)
- wt and mutant ferrous iron transporter LIT1 in *L. amazonensis* (Jacques et al. 2010)
- membrane acid phosphatase in *L. donovani* (Papadaki et al. 2015)



- functional characterization of parasite proteins
- investigation of parasite-host interactions
- *in vivo* and *in vitro* screening of anti-leishmanial drugs
- diagnostics and vaccines development

Collaborations

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References LEXSY applications for parasitologists

Fluorescent/luminiscent cells

- DsRed in *L. major* (Chagas *et al.* 2014, Rabhi *et al.* 2016)
- near-infrared *L. infantum* strains (Calvo-Álvarez *et al.* 2015, Balaña-Fouce *et al.* 2012)
- EGFP-LUC in *L. major* (Taheri *et al.* 2015)
- LUC bioluminescent *L. donovani* (García-Hernández *et al.* 2015)
- LUC bioluminescent *L. amazonensis* (Lang *et al.* 2005)
- eGFP and RFP in *L. donovani* (Sadlova *et al.* 2011)
- EGFP in *L. major* (Bolhassani *et al.* 2011, Gonzalez-Leal *et al.* 2014)
- EGFP in *L. donovani* (Barak *et al.* 2005)
- mCherry in *L. major* and *L. donovani* (Vacchini *et al.* 2014, Calvo-Alvarez *et al.* 2012)

References LEXSY applications for parasitologists

Antigens

- *T. brucei gambiense* surface ags ISG65, VSG LiTat 1.3 and VSG LiTat 1.5 (Rooney *et al.* 2015)
- Surface antigen PSA-38S *L. amazonensis* (Chamakh-Ayari *et al.* 2014)
- sand fly salivary antigen PpSP15-EGFP in *L. tarentolae* (Katebi *et al.* 2015)
- Polytope -PT-GFP (Seyed *et al.* 2015)
- Cysteine proteases cpa-cpb fusions (Taheri *et al.* 2014, Zahedifard *et al.* 2014, Saljoughian *et al.* 2013)

Functional studies

- membrane acid phosphatase in *L. donovani*, *LdMACP* (Papadaki *et al.* 2015)
- histone acetyltransferase HAT3 in *L. donovani* (Kumar *et al.* 2015)
- MAP kinase MPK4 in *L. major* (Dacher *et al.* 2014)
- Casein Kinase 1 in *L. donovani* (Rachidi *et al.* 2014)
- apurinic/apirimidinic endonuclease AP2 *T. cruzi* in *L. tarentolae* (Sepúlveda *et al.* 2014)
- LPG3 of *L.infantum* in *L. tarentolae* (Pirdel *et al.* 2012)
- *L. donovani* actin (LdACT) in *L. tarentolae* (Kumar *et al.* 2012)